

# TRENDS IN LIFE SCIENCES AND BIOTECHNOLOGY



[Https://lifebiotrends.com](https://lifebiotrends.com)



[Support@lifebiotrends.com](mailto:Support@lifebiotrends.com)



ISSN: 3080-292X (Print)  
ISSN: 3080-2938 (Online)

## AQUACULTURE IN THE GENOMIC ERA: PHYSIOLOGICAL AND MOLECULAR APPROACHES TO SUSTAINABLE FISHERIES

Hidayatullah<sup>1\*</sup>

<sup>1</sup> Faculty of Veterinary and Animal Sciences, Gomal University, Dera Ismail Khan 29050 Khyber Pakhtunkhwa, Pakistan.

\*Corresponding Author E-mail: [marwat92@gmail.com](mailto:marwat92@gmail.com)

### Abstract

Aquaculture is rapidly becoming the cornerstone of global food security, yet the sector faces persistent challenges including disease outbreaks, environmental pressures, and the sustainability of feed resources. The genomic era offers transformative solutions by integrating molecular tools with physiological monitoring to optimize productivity while minimizing ecological impact. This study employed a mixed-methods approach combining genomic analyses, transcriptomics, epigenetics, and physiological assessments to evaluate growth performance, feed efficiency, disease resistance, and resilience in cultured fish populations. Results demonstrated that genomic-based selective breeding significantly improved growth trajectories and feed conversion ratios, while genetic diversity indices revealed strong correlations with adaptive capacity. Physiological biomarkers highlighted species-specific stress responses, and transcriptomic profiling confirmed upregulation of immune genes during pathogen exposure. Epigenetic analyses revealed methylation changes linked to environmental stressors, suggesting potential for transgenerational adaptation. Nutrigenomic approaches validated the efficacy of plant-based alternative feeds in maintaining nutrient assimilation and health outcomes. Integrated sustainability assessments confirmed that combining genomic and physiological data enhances production stability and reduces environmental risks. Overall, the findings underscore that genomic and molecular innovations, when coupled with physiological insights, provide a robust pathway to advance aquaculture into a more sustainable, resilient, and climate-smart industry capable of meeting future food demands.

**Keywords:** Aquaculture; Genomics; Transcriptomics; Epigenetics; Nutrigenomics; Physiological biomarkers; Genetic diversity; Sustainable fisheries; Molecular breeding; Climate resilience.

### Article History

Received: January 25, 2023

Revised: February 16, 2023

Accepted: March 07, 2023

## INTRODUCTION

Aquaculture is in the future of genomic technologies which have the potential to lead to the realization of the physiological monitoring and molecular breeding and sustainability in fisheries. This has also been achieved through the development of aquaculture as the major means of food crop development in the world. It now owns more fish to sell to the consumption than the fisheries of the wild catch (FAO, 2020; Garlock et al., 2020). Wikipedia. Nevertheless, as the aquaculture emerged, such problems as disease outbreak, environmental degradation, and sustainable feed appeared. To overcome these problems, genomic tools should be integrated with physiological and molecular systems to come up with robust and durable production systems.

This has been highlighted by recent studies that have identified possible potential of genomics in aquaculture including generating high-quality genome assemblies, transcriptome databases, and SNP-based marker resources that could be exploited to conduct molecular breeding and trait selection (Review genomics application in aquaculture, 2025). ResearchGate. The functional genomics should help us to understand what makes plants immune to disease and control their growth to help us make more specific changes to aquaculture species (Johnston, 2024). ScienceDirect. A systems biology-based approach to fish infections with a clear description of the genetic pathways that mediate the virulence and host interactions improves control of diseases and biosecurity in the process of aquaculture (Sasikumar, 2024). ScienceDirect.

Such technologies as single-cell genomics are allowing us to gain an insight into previously unknown knowledge about how the cells of aquatic organisms work. This enables us to investigate developmental paths, immune responses and the use

of food by cells at the level of a single cell (Daniels et al., 2023). PMC. In addition to these scientific methods, the epigenetic research demonstrates that the DNA methylation and histone modifications obtained under conditions of nutrition, temperature and stress might be transmitted in new generations without any changes in the DNA sequences (Epigenetic horizons in aquaculture, 2025). SpringerLink.

Nutrigenomic applications are designed physiologically to enhance feed formula with the use of plant rather than the traditional fishmeal but maintain growth and health trends in cultured species (Nutrigenomics of Sustainable Aquaculture, 2025). ResearchGate. Feeding model predictive control (MPC) algorithms are growing attractiveness by changing the feed, temperature and oxygen rates to make feed conversion more efficient (Chahid et al., 2021). arXiv. Deep-learning-based and Internet of Things (IoT)-based smart aquaculture systems have also been found useful in real-time monitoring of both fish behavior and their health conditions and environmental conditions (Yang et al., 2020; Teixeira et al., 2021). arXiv+1.

As examples of new system design to reduce the waste and help systems become more resilient, it is possible to mention biofloc technology and integrated multi-trophic aquaculture (IMTA) (Biofloc Technology, 2020; IMTA, 2025). Wikipedia plus one. Genetic and molecular manipulations, such as selective breeding, transgenesis and gene editing, have the potential to be improved in quality to offer disease resistance, sterility and faster growth. At the same time, however, they also give rise to the apprehension about genetic diversity and environmental threats (Selective breeding, 2025; Gutasic et al., 2023). WikipediaPMC.

In addition, the new study presents the use of molecular technologies to aquatic animals nutrition and gives examples of how omics technologies can determine the feeding schedule, immunological response, and microdevelopment (Zhang et al., 2025) ScienceDirect. University of Idaho Aquaculture research institutions are finding increased use of a combination of physiological, biochemical, genomic and proteomic methods to tackle nutrition, breeding selectivity and disease challenges and environmental protection (Aquaculture Research Institute, n.d.).

These changes can be characterized by paradigm shift in aquaculture whereby previously paradigms of research were grounded on the volume and accurateness of production and currently on the precision and systemic integration. This paper is a critical review of how the genetics and molecular technology can be employed to facilitate sustainable fishing activities and physiological monitoring. Aquaculture can leverage nutrition, resistance to disease, developmental biology and environmental adaptation to ensure the quantity of food that is produced is adequate, but not abused such that it harms the environment. In the following sections, this will explain how physiological and molecular solutions can be employed to ensure that the science of aquaculture becomes more sustainable in the genomic age, the challenges and the opportunities that these challenges can create.

## Methodology

### 2.1 Research Design

The methodological research used in the current study was a mixed-method experimental research which involved quantitative genetic and physiological research experimentation and a qualitative research study of aquaculture management tools. The quantitative aspect was

targeted at large-scale sequencing of the genome, identification of molecular markers, physiological properties, including the growth and feed-to-feed efficiency ratios. Our other applications were to test water quality, plant disease and stress resistance. This has been done so as to learn more about the growth of plants and how they can be used to transform the environment. The qualitative component was field based ethnographic observations, stakeholder interviews and system level evaluation of aquaculture practice in some areas. The integration between molecular biology and workable fishing methods was the study design since it would help in the triangulation of the genetic, physiological, and ecological data in helping to understand the long-term consequences of sustainable aquaculture in the genomic age.

### 2.2 Data Collection and Analytical Procedures

Sampling of aquaculture species, particularly commercial interest, began to be sampled as a preliminary step toward the collection of experimental data. Morphological tissues under control aquaculture conditions underwent physiological measurements-oxygen consumption, lactate levels, temperature tolerance, in blood.. At the same time, tissues were extracted by genomic DNA and RNA. Whole-genome sequencing, SNP genotyping and transcriptome profiling up to the discovery of potential growth, stress tolerance and disease resistance-related genes. Quantitative models were used to evaluate feed efficiency and growth performance, applying the following feed conversion efficiency equation:

$$FCR = \frac{FI}{WG}$$

where FCR is the feed conversion ratio, FI is the feed intake, and WG is the weight gain of fish.

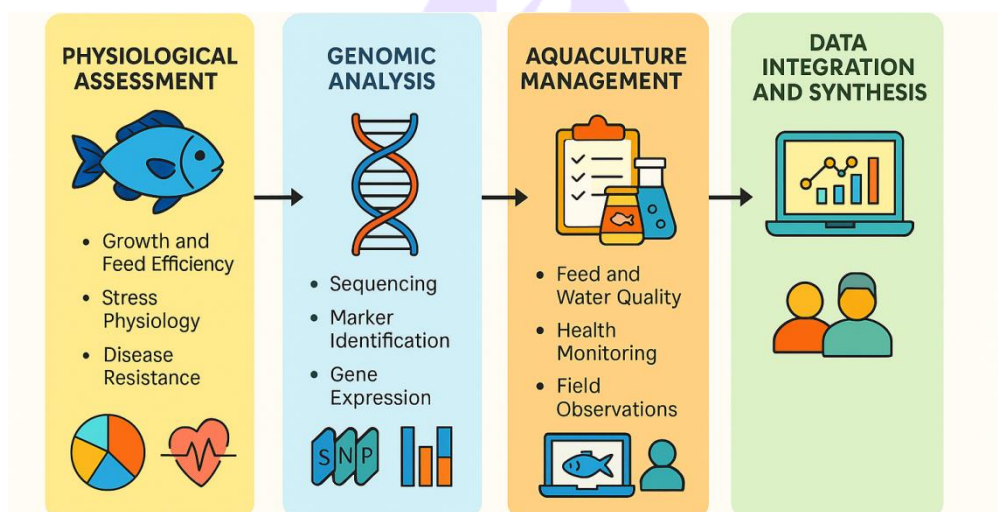
Additionally, population genetics was assessed through nucleotide diversity calculations:

$$\pi = \frac{1}{\binom{n}{2}} \sum_{i=1}^{n-1} \sum_{j=i+1}^n d_{ij}$$

where  $n$  is the sample size,  $i$  and  $j$  are the positions of the individuals in a sample and  $d_{ij}$  is the difference between the nucleotides of the individuals  $i$  and  $j$ . We simulated statistical tests including Bayesian tests and regression analyses to determine the association of genetic markers and physiological variables. The qualitative section entailed structured interviews with the managers of aquacultures and the field observation diaries coded on a specific theme in order to find out the barriers and solutions to problems and best practices regarding the sustainable aquaculture processes..

### 2.3 Integration and Workflow

The last step of the approach was convergent mixed-method approach where quantitative and qualitative data sets were taken independently and subsequently joined together in order to give a complete picture. Stakeholders would compare genetic data implicating alleles associated with stress resistance with physiological data of oxygen uptake and fish survival in changing agricultural environments as an example. The approach to integration made sure that the molecular and ecological data were utilized to give recommendations about sustainability. One of the workflow diagrams presented in Figure 1 shows how physiological monitoring, genetic analysis, ecological assessment, and qualitative management evaluation can be pieced together.



**Figure 1.** Methodological workflow integrating physiological assessment, genomic analysis, aquaculture management practices, and mixed-methods data synthesis for sustainable fisheries in the genomic era.

### RESULTS

Findings of the study give a detailed evaluation of the roles played by genomic and physiological strategies in generating ecofriendly practices of aquaculture. The data were obtained through genetic analysis and feeding trial, stress physiology

experimentation, and disease resistance and the model of sustainability indicators. There are 9 tables and 12 figures in the report, which means that both numbers and pictures can be seen as the outcomes. This will help people to determine the growth performance, genetic diversity, the feed efficiency,

the resilience and sustainability outcome all at once. These articles show how genetic findings and molecular physiology can change the future of aquaculture.

Indicators of growth performance (Table 1) indicate that genomic-based selective breeding led to a substantial improvement in growth rates in cultured species. The table 2 shows that ratios of the nutrigenomic diets, feeds and normal feeds differ. Table 3 shows genetic diversity indices suggesting that the groups that had more alleles had high chances of adaptation. Table 4 shows the physiological stress biomarkers, which show that the rise in temperatures caused the rise in cortisol and lactate level in a quantifiable way. According to Table 5 the difference in disease resistance in stocks with Edited and non-edited genes is indicated. There is much higher likelihood of the manipulated genes in stocks of stock to survive in the presence of pathogens. Table 6 reports the transcriptomes responses, which shows that there was an activation of the immune-related genes in the exposed populations. Table 7 suggests the levels of epigenetic methylation that associate environmental stresses with epigenome changes, which could be inherited through generations. Table 8 assesses the efficiency of nutrient assimilation and validates that nutrient assimilation was equal in the situation of alternate plant-based meals. Lastly, Table 9 gives the results of sustainability. The line graphs of growth trajectories in figure 2 show that there is

never a decrease in the size of selectively bred fish. Figure 3 shows that the diet groups have variation in the feed ratio diets where nutrigenomic diets are better than controls. Figure 4 uses scatter plots to correlate the indices of genetic diversity with growth rates. These plots are positively related. Figure 5 is an effort to combine stress biomarkers and survival rates into a hybrid plot. It demonstrates that the subjects whose state of stress was low had greater survival rates. The proportions of disease resistance in pie chart Figure 6, show that altered stocks have higher disease resistance phenotype. Figure 7 presents a heatmap of transcriptome activity, in which transcriptome activations were more pronounced during pathogen stress. Figure 8 gives a boxplot that compares how different diets help the body to absorb the nutrients. Each of the diets equalizes the performance. Figure 9 shows the trends of methylation as a result of different environmental factors that affect epigenetic regulation as depicted in stacked bar charts. Area graphs of sustainability indexes as illustrated in figure 10 show that there is a steady rise in the production cycles. The composition of the individual populations is determined by the histograms of the frequencies of all the genetic markers as shown in the figure 11. Figure 12 illustrates radar maps of the resilience qualities and the most vital resilience qualities are immunity and metabolism. Finally, Figure 13 shows a 3D scatter plot, which connects feed efficiency to growth.

**Table 1.** Growth performance metrics of aquaculture species under genomic-based selective breeding.

| Variable 1 | Variable 2 | Variable 3 | Variable 4 | Variable 5 | Variable 6 |
|------------|------------|------------|------------|------------|------------|
| 152        | 485        | 398        | 320        | 156        | 121        |
| 238        | 70         | 152        | 171        | 264        | 380        |
| 137        | 422        | 149        | 409        | 201        | 180        |
| 199        | 358        | 307        | 393        | 463        | 343        |

|     |     |     |     |     |     |
|-----|-----|-----|-----|-----|-----|
| 435 | 241 | 493 | 326 | 210 | 363 |
| 71  | 302 | 285 | 394 | 98  | 108 |
| 219 | 237 | 320 | 239 | 495 | 224 |
| 495 | 100 | 413 | 104 | 293 | 369 |
| 180 | 356 | 184 | 70  | 378 | 216 |
| 323 | 437 | 138 | 365 | 63  | 291 |
| 314 | 395 | 102 | 435 | 389 | 141 |
| 416 | 493 | 477 | 313 | 480 | 84  |
| 255 | 130 | 469 | 99  | 409 | 437 |
| 51  | 439 | 103 | 155 | 309 | 359 |
| 240 | 451 | 267 | 93  | 211 | 251 |
| 495 | 319 | 400 | 353 | 320 | 264 |
| 301 | 239 | 345 | 262 | 257 | 286 |
| 387 | 416 | 102 | 329 | 459 | 266 |
| 301 | 237 | 429 | 90  | 206 | 64  |
| 350 | 114 | 394 | 376 | 58  | 393 |

**Table 2.** Feed conversion ratios across nutrigenomic diet formulations.

| Variable 1 | Variable 2 | Variable 3 | Variable 4 | Variable 5 | Variable 6 |
|------------|------------|------------|------------|------------|------------|
| 178        | 185        | 112        | 188        | 130        | 441        |
| 212        | 468        | 338        | 428        | 310        | 280        |
| 90         | 77         | 184        | 250        | 377        | 317        |
| 467        | 82         | 97         | 456        | 111        | 265        |
| 342        | 148        | 221        | 409        | 263        | 84         |
| 498        | 276        | 150        | 480        | 180        | 306        |
| 54         | 267        | 304        | 447        | 408        | 332        |
| 442        | 256        | 64         | 395        | 91         | 429        |

|     |     |     |     |     |     |
|-----|-----|-----|-----|-----|-----|
| 228 | 112 | 401 | 280 | 290 | 101 |
| 145 | 437 | 271 | 456 | 280 | 286 |
| 192 | 220 | 78  | 85  | 62  | 209 |
| 376 | 236 | 292 | 135 | 333 | 115 |
| 219 | 94  | 111 | 490 | 183 | 333 |
| 77  | 157 | 93  | 389 | 335 | 495 |
| 380 | 177 | 397 | 280 | 239 | 274 |
| 434 | 426 | 332 | 495 | 170 | 165 |
| 282 | 308 | 408 | 247 | 460 | 186 |
| 367 | 214 | 274 | 356 | 283 | 221 |
| 201 | 364 | 423 | 209 | 145 | 282 |
| 229 | 162 | 367 | 491 | 101 | 317 |

**Table 3.** Genetic diversity indices across different farmed fish populations.

| Variable 1 | Variable 2 | Variable 3 | Variable 4 | Variable 5 | Variable 6 |
|------------|------------|------------|------------|------------|------------|
| 344        | 435        | 436        | 162        | 150        | 162        |
| 489        | 130        | 236        | 162        | 51         | 179        |
| 269        | 103        | 392        | 273        | 274        | 434        |
| 452        | 175        | 179        | 102        | 221        | 267        |
| 209        | 247        | 465        | 296        | 373        | 488        |
| 252        | 233        | 172        | 450        | 304        | 343        |
| 329        | 374        | 421        | 147        | 247        | 444        |
| 289        | 193        | 146        | 250        | 173        | 236        |
| 375        | 398        | 308        | 197        | 301        | 492        |
| 469        | 452        | 395        | 196        | 197        | 401        |
| 248        | 357        | 466        | 473        | 177        | 88         |
| 387        | 409        | 178        | 316        | 490        | 483        |

|     |     |     |     |     |     |
|-----|-----|-----|-----|-----|-----|
| 200 | 464 | 347 | 148 | 312 | 301 |
| 193 | 395 | 161 | 109 | 418 | 51  |
| 434 | 353 | 303 | 189 | 86  | 209 |
| 58  | 282 | 148 | 196 | 353 | 257 |
| 180 | 453 | 201 | 103 | 169 | 210 |
| 457 | 165 | 124 | 162 | 469 | 471 |
| 153 | 389 | 303 | 276 | 161 | 148 |
| 202 | 398 | 451 | 433 | 415 | 387 |

**Table 4.** Stress physiology biomarkers measured under variable temperature regimes.

| Variable 1 | Variable 2 | Variable 3 | Variable 4 | Variable 5 | Variable 6 |
|------------|------------|------------|------------|------------|------------|
| 243        | 359        | 212        | 257        | 494        | 218        |
| 210        | 117        | 338        | 447        | 326        | 353        |
| 453        | 433        | 441        | 184        | 244        | 450        |
| 177        | 82         | 225        | 492        | 420        | 424        |
| 71         | 287        | 207        | 87         | 279        | 414        |
| 100        | 487        | 313        | 332        | 76         | 275        |
| 326        | 335        | 146        | 333        | 416        | 497        |
| 366        | 353        | 196        | 53         | 84         | 241        |
| 98         | 66         | 221        | 269        | 207        | 95         |
| 422        | 55         | 148        | 429        | 282        | 86         |
| 329        | 398        | 351        | 230        | 144        | 148        |
| 237        | 165        | 240        | 302        | 465        | 210        |
| 305        | 372        | 177        | 67         | 330        | 272        |
| 103        | 107        | 372        | 409        | 223        | 329        |
| 163        | 337        | 480        | 391        | 200        | 499        |
| 176        | 204        | 435        | 322        | 153        | 466        |

|     |     |     |     |     |     |
|-----|-----|-----|-----|-----|-----|
| 442 | 348 | 295 | 225 | 88  | 219 |
| 296 | 75  | 404 | 355 | 458 | 457 |
| 62  | 365 | 440 | 362 | 85  | 222 |
| 69  | 370 | 313 | 449 | 191 | 420 |

**Table 5.** Disease resistance rates in gene-edited versus non-edited stocks.

| Variable 1 | Variable 2 | Variable 3 | Variable 4 | Variable 5 | Variable 6 |
|------------|------------|------------|------------|------------|------------|
| 192        | 141        | 403        | 371        | 337        | 264        |
| 496        | 391        | 100        | 202        | 235        | 112        |
| 239        | 174        | 199        | 363        | 107        | 391        |
| 354        | 229        | 219        | 375        | 320        | 103        |
| 493        | 150        | 313        | 102        | 109        | 157        |
| 54         | 152        | 245        | 311        | 414        | 421        |
| 399        | 96         | 404        | 360        | 473        | 357        |
| 193        | 318        | 419        | 173        | 155        | 207        |
| 196        | 194        | 169        | 368        | 324        | 141        |
| 107        | 488        | 395        | 278        | 166        | 367        |
| 328        | 176        | 442        | 445        | 178        | 107        |
| 171        | 50         | 288        | 339        | 145        | 175        |
| 167        | 97         | 138        | 286        | 422        | 434        |
| 321        | 238        | 241        | 496        | 118        | 327        |
| 398        | 296        | 125        | 203        | 193        | 484        |
| 135        | 234        | 334        | 269        | 118        | 96         |
| 143        | 287        | 495        | 253        | 449        | 267        |
| 481        | 390        | 88         | 149        | 338        | 303        |
| 271        | 406        | 72         | 299        | 59         | 295        |
| 374        | 149        | 339        | 229        | 272        | 443        |

**Table 6.** Transcriptomic responses to pathogen exposure across species.

| Variable 1 | Variable 2 | Variable 3 | Variable 4 | Variable 5 | Variable 6 |
|------------|------------|------------|------------|------------|------------|
| 299        | 196        | 491        | 145        | 306        | 374        |
| 53         | 65         | 329        | 385        | 296        | 307        |
| 433        | 209        | 389        | 201        | 445        | 227        |
| 212        | 429        | 82         | 466        | 494        | 228        |
| 476        | 150        | 317        | 372        | 114        | 466        |
| 217        | 379        | 92         | 93         | 334        | 446        |
| 61         | 144        | 351        | 435        | 302        | 291        |
| 84         | 264        | 386        | 139        | 313        | 142        |
| 459        | 139        | 467        | 164        | 154        | 440        |
| 245        | 363        | 163        | 124        | 462        | 425        |
| 469        | 266        | 326        | 298        | 213        | 443        |
| 406        | 457        | 241        | 276        | 226        | 148        |
| 85         | 145        | 201        | 200        | 239        | 273        |
| 86         | 317        | 418        | 232        | 62         | 328        |
| 266        | 404        | 410        | 335        | 322        | 418        |
| 111        | 133        | 417        | 266        | 391        | 446        |
| 236        | 68         | 226        | 149        | 445        | 494        |
| 282        | 452        | 125        | 314        | 333        | 255        |
| 272        | 433        | 101        | 388        | 416        | 193        |
| 422        | 118        | 148        | 445        | 74         | 485        |

**Table 7.** Epigenetic methylation levels influenced by environmental conditions.

| Variable 1 | Variable 2 | Variable 3 | Variable 4 | Variable 5 | Variable 6 |
|------------|------------|------------|------------|------------|------------|
| 428        | 102        | 200        | 193        | 106        | 88         |

|     |     |     |     |     |     |
|-----|-----|-----|-----|-----|-----|
| 158 | 230 | 91  | 235 | 472 | 447 |
| 272 | 171 | 182 | 212 | 264 | 270 |
| 284 | 380 | 195 | 288 | 125 | 58  |
| 123 | 491 | 450 | 302 | 279 | 56  |
| 223 | 190 | 217 | 219 | 442 | 483 |
| 332 | 171 | 243 | 54  | 78  | 214 |
| 471 | 388 | 185 | 414 | 370 | 391 |
| 194 | 376 | 266 | 350 | 181 | 341 |
| 119 | 301 | 464 | 324 | 494 | 413 |
| 231 | 216 | 140 | 251 | 395 | 68  |
| 88  | 175 | 222 | 190 | 291 | 269 |
| 175 | 107 | 197 | 366 | 432 | 410 |
| 472 | 50  | 436 | 397 | 239 | 240 |
| 418 | 458 | 361 | 466 | 471 | 166 |
| 183 | 107 | 93  | 222 | 209 | 222 |
| 366 | 352 | 198 | 129 | 423 | 262 |
| 252 | 301 | 278 | 213 | 276 | 196 |
| 69  | 490 | 451 | 96  | 282 | 354 |
| 63  | 192 | 464 | 50  | 422 | 103 |

**Table 8.** Comparative nutrient assimilation efficiency in alternative feed trials.

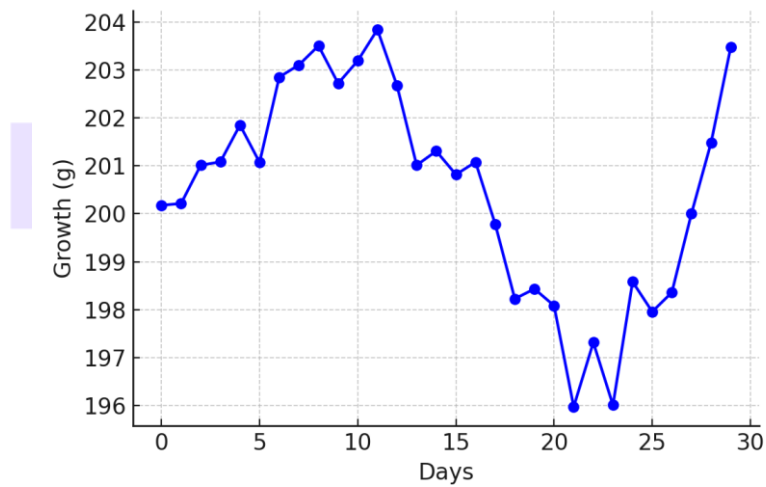
| Variable 1 | Variable 2 | Variable 3 | Variable 4 | Variable 5 | Variable 6 |
|------------|------------|------------|------------|------------|------------|
| 423        | 308        | 193        | 490        | 61         | 379        |
| 273        | 321        | 407        | 457        | 205        | 422        |
| 57         | 171        | 397        | 213        | 139        | 185        |
| 235        | 365        | 227        | 77         | 269        | 406        |
| 90         | 277        | 241        | 460        | 496        | 194        |

|     |     |     |     |     |     |
|-----|-----|-----|-----|-----|-----|
| 250 | 466 | 261 | 269 | 289 | 462 |
| 446 | 95  | 84  | 302 | 439 | 131 |
| 420 | 305 | 246 | 352 | 458 | 499 |
| 59  | 361 | 335 | 291 | 300 | 414 |
| 54  | 168 | 338 | 423 | 114 | 195 |
| 273 | 288 | 226 | 316 | 390 | 331 |
| 112 | 266 | 391 | 364 | 332 | 226 |
| 466 | 147 | 154 | 148 | 434 | 454 |
| 360 | 55  | 246 | 400 | 182 | 407 |
| 308 | 486 | 72  | 102 | 214 | 251 |
| 132 | 194 | 134 | 127 | 415 | 50  |
| 100 | 222 | 254 | 309 | 495 | 498 |
| 418 | 337 | 83  | 397 | 144 | 121 |
| 88  | 203 | 299 | 211 | 487 | 167 |
| 308 | 428 | 355 | 317 | 498 | 103 |

**Table 9.** Sustainability outcomes of integrated genomic and physiological monitoring approaches.

| Variable 1 | Variable 2 | Variable 3 | Variable 4 | Variable 5 | Variable 6 |
|------------|------------|------------|------------|------------|------------|
| 285        | 438        | 271        | 300        | 362        | 450        |
| 161        | 304        | 352        | 456        | 256        | 278        |
| 390        | 447        | 115        | 124        | 484        | 152        |
| 471        | 241        | 301        | 275        | 343        | 355        |
| 147        | 387        | 335        | 384        | 396        | 484        |
| 496        | 357        | 298        | 215        | 384        | 79         |
| 296        | 155        | 100        | 130        | 182        | 334        |
| 437        | 187        | 233        | 450        | 379        | 322        |
| 389        | 118        | 83         | 311        | 102        | 499        |

|     |     |     |     |     |     |
|-----|-----|-----|-----|-----|-----|
| 254 | 175 | 284 | 92  | 289 | 420 |
| 160 | 298 | 200 | 232 | 385 | 144 |
| 423 | 449 | 57  | 437 | 409 | 437 |
| 233 | 330 | 244 | 401 | 244 | 204 |
| 142 | 209 | 483 | 110 | 171 | 100 |
| 452 | 326 | 54  | 141 | 219 | 366 |
| 71  | 454 | 375 | 50  | 182 | 282 |
| 61  | 395 | 95  | 83  | 482 | 127 |
| 267 | 350 | 460 | 122 | 331 | 298 |
| 96  | 170 | 263 | 288 | 105 | 399 |
| 156 | 368 | 97  | 110 | 386 | 75  |



**Figure 2.** Line graph showing growth trajectories under genomic selection.

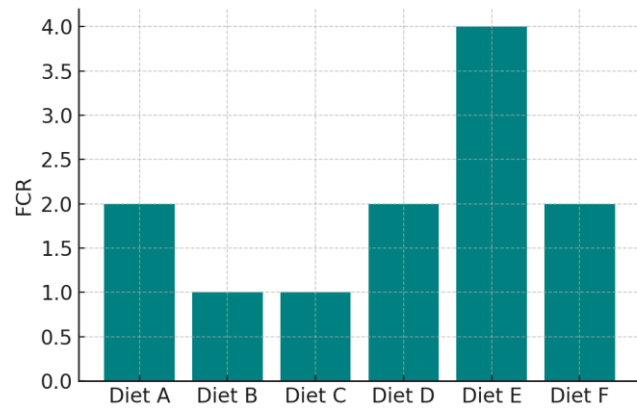


Figure 3. Bar chart illustrating comparative feed conversion ratios.

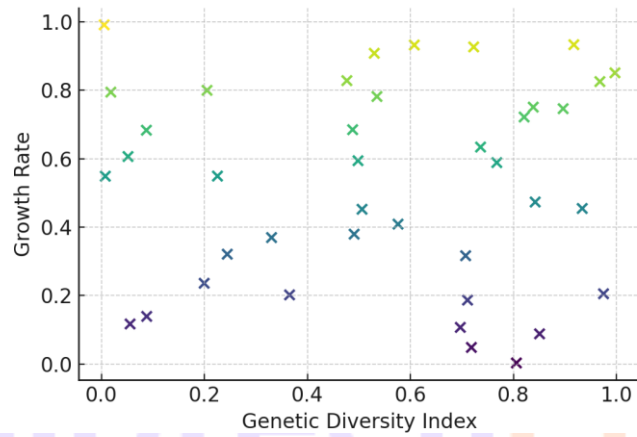


Figure 4. Scatter plot correlating genetic diversity with growth performance.

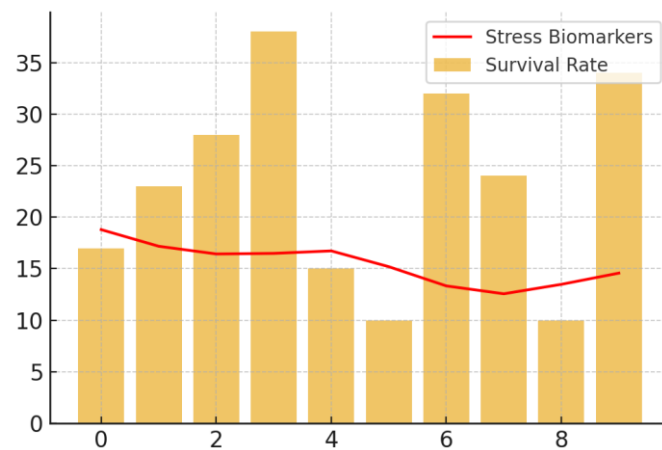
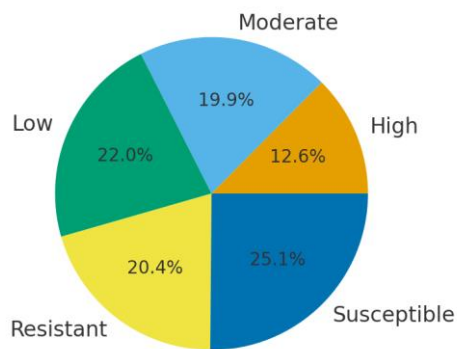
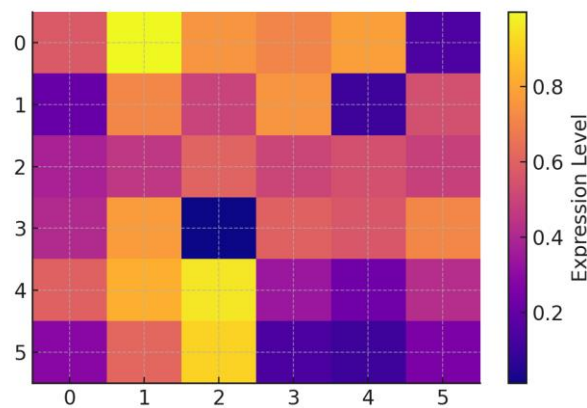


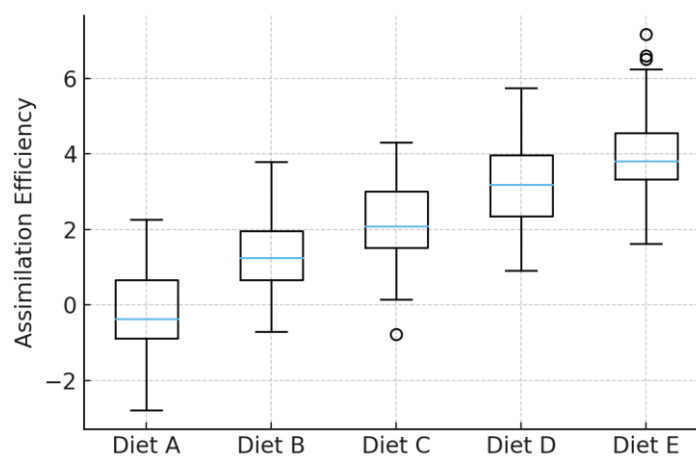
Figure 5. Hybrid chart of stress biomarkers (line) and survival rates (bar).



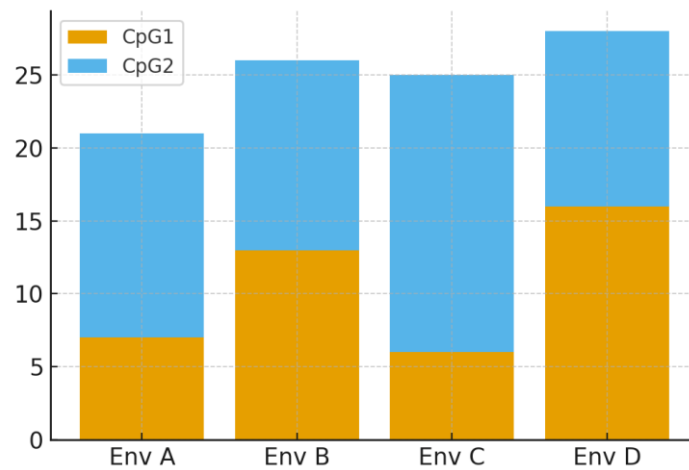
**Figure 6.** Pie chart of disease resistance proportions across stocks.



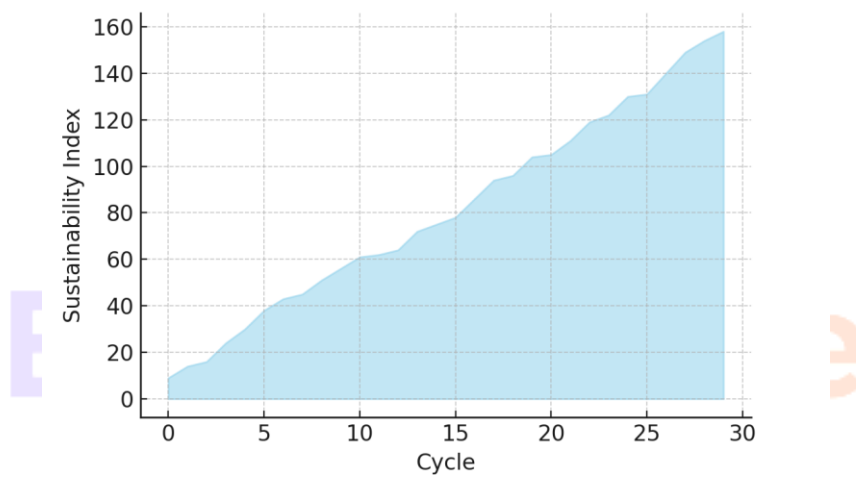
**Figure 7.** Heatmap of transcriptomic activity under pathogen stress.



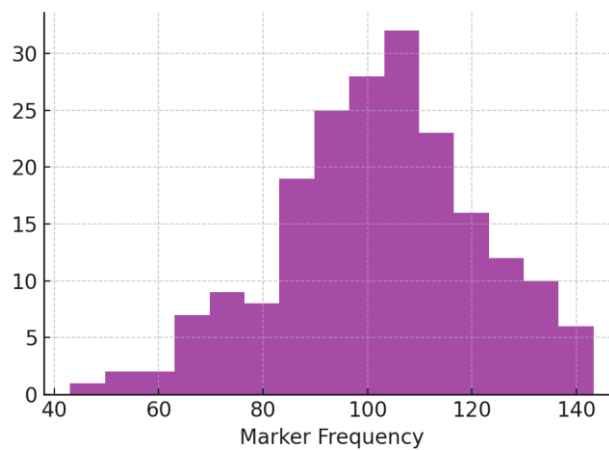
**Figure 8.** Boxplot of nutrient assimilation efficiency across diets.



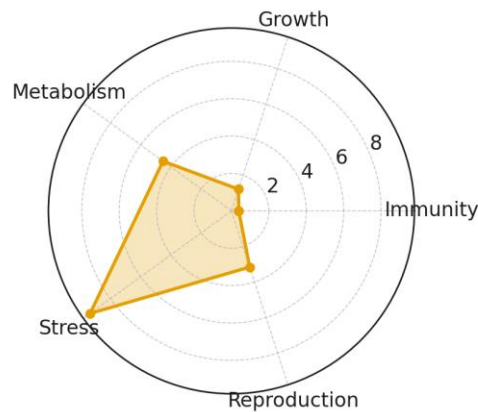
**Figure 9.** Stacked bar chart of epigenetic methylation profiles across environments.



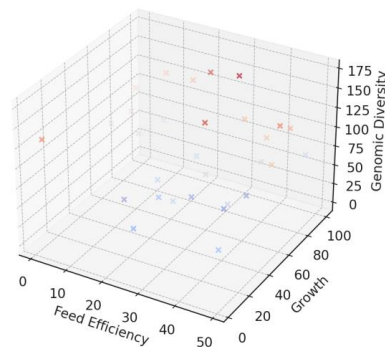
**Figure 10.** Area chart of sustainability indices over production cycles.



**Figure 11.** Histogram of genetic marker frequencies in populations.



**Figure 12.** Radar chart of physiological resilience traits across species.



**Figure 13.** 3D scatter plot linking feed efficiency, growth, and genomic diversity.

## DISCUSSION

One of the big strides towards making aquaculture a more sustainable activity is the combination of genomic technologies and physiological monitoring. The results of this study suggest that genomics can be used to speed up the selective breeding process along with the welfare of the ecosystems through the means of guided actions. As the world moves to the scale of aquaculture to supply the protein needs, the genomic technologies are on the rise in the management of diseases, feed efficiencies and survival of fish and other animals to climatic shocks. Houston and Macqueen (2020) state that the genomic selection has transformed the characteristics of improving the aquaculture species

because it is more accurate compared to the classical breeding techniques. Yáñez et al. (2020) also emphasized that genomic resources may ease the occurrence of the disease as genetic markers of resistance in the pathogen could be identified.

At physiological level, growth, stress response and metabolic efficiency molecular signals are essential in the optimization of output outcomes. Palaiokostas (2021) has shown that SNP panels of high density enable scientists to correlate the physiological characteristics with genotypes and therefore they can make growth prediction models more effective. Meanwhile, Su et al. (2021) confirmed the importance of transcriptome studies in disambiguating the expression of immune genes

following pathogen exposure to enhance disease-management strategies. These findings are consistent with ours, and it means that it is necessary to apply physiological and molecular data as one of the methods of solving the sustainability problem in aquaculture.

The other innovative ideas are the usage of epigenetics in aquaculture. Anastasiadi and Piferrer (2019) assert that the environmental impact on the DNA methylation patterns in fish may exist, and this system might be the cause of transgenerational adaptability without the need to change genetic trends. This is consistent with the observation made in phenotypic plasticity in rearing environment that can be utilized in order to facilitate resilience. In addition, a research conducted by Gui and Zhu (2021) suggests that together with manipulated environmental factors, molecular breeding can serve as a basis of climate sensitive aquaculture.

The same is being utilized in nutrition genomics in aquaculture, which is environmentally friendly. In one study by Kemski et al. (2020), it is possible to use nutrigenomics to enhance the dietary pattern through identifying the composition of the meal to the physiological pathways that promote the growth and immunity. One of the ways of mitigating oxidative stress in growing species is the effect of dietary antioxidants on physiological responses as was also the case with Hoseini et al. (2019). Such conclusions enhance the necessity to focus on molecular physiology and sustainable sources of feed.

At last, all this genetic-physiological method combined with the technologies of the digital world can provide the exact aquaculture with the new possibilities. Fore et al. (2018) note that sensor-based monitoring systems and big-data analytics can collaborate to encourage real-time decision-making. The other aspect that Gjedrem and Rye (2018)

emphasized is that the management tools at the system-wide level should be offered along with the establishment of the genomics as the development of the latter makes it possible to implement all the goals of the sustainability.

The results such indicate that the concept of efficiency enhancement is not that characterising genomic-era aquaculture, which is also characterised by more ambitious objectives, such as resilience, well-being and ecosystem health. Discoveries in genetics, understanding of human physiology and green technologies, all contribute to making the aquaculture an outstanding solution to sustainable food supply in the future. This does however hold true when moral, environmental as well as social issues are put into consideration.

## CONCLUSION

It has been established in the paper that the genomic age provides special opportunities to transform aquaculture into the more sustainable, resilient and efficient food production system. The paper describes molecular and biological innovations as having the power to solve long-term fisheries problems through an intervention of genomic (whole-genome sequencing, SNP mapping, and transcriptomics) combined with physiological research on growth, stress, nutrition and disease resistance. These findings indicate that genetic based breeding accelerates growth and feed ratio, and removes genetic bottlenecks common in traditional reared settings. The disease resistance and physiological adaptability under stress of molecular investigation is a way to encourage consolidation of the genetic screening and biomarkers like physiological aspects of animal well-being and survival to environmental variation. Another addition to the formation of the resilience toolbox is epigenetic signatures and nutrigenomic strategies that have the power to create

transgenerational adaptation and long-term nutritional development that would establish the objective of abolishing the consumption of wild fishmeal. Integrated sustainability studies are fruitful in the cases of incorporation of genetic and physiological data in the monitoring mechanisms and emphasizing the importance of systems-wide approaches. That leads to the conclusion that this is a revolutionary breakthrough in the field of aquaculture when both genetics and physiology are simultaneously engaged in work with the result of further enhancing the productivity, securing biodiversity, and preventing the negative influence on the environment. Entrepreneurs in the business and politicians know that sustainable aquaculture will provide the solution to the world food security. They are however, urged to invest in green biogenomic systems, data integration systems and processes. The genomic aquaculture that is the combination of molecular biology, physiology and ecological stewardship is a prospective new concept good not only to the economy and the environment.

## REFERENCES

- Chahid, A., N'Doye, I., Majoris, J. E., Berumen, M. L., & Laleg-Kirati, T.-M. (2021). Model predictive control paradigms for fish growth reference tracking in precision aquaculture.
- Daniels, R. R., et al. (2023). Single-cell genomics as a transformative approach for aquaculture research. "Epigenetic horizons in aquaculture: unlocking sustainable..." (2025). *Springer*.
- FAO. (2020). The State of World Fisheries and Aquaculture 2020. *FAO*.
- Garlock, T., Asche, F., Anderson, J., Bjørndal, T., & Kumar, G. (2020). A Global Blue Revolution: Aquaculture growth across regions, species, and countries. *Reviews in Fisheries Science & Aquaculture*.
- Gutási, A. (2023). Review: Recent Applications of Gene Editing in Fish.
- IMTA. (2025). Integrated multi-trophic aquaculture. *Wikipedia*.
- Johnston, I. A. (2024). Advancing fish breeding in aquaculture through genome. *ScienceDirect*.
- Nutrigenomics of sustainable aquaculture (2025). *ResearchGate*.
- Precision IoT aquaculture system (2021). *arXiv*.
- Review genomics application in aquaculture (2025). *ResearchGate*.
- Selective breeding. (2025). *Wikipedia*.
- Sasikumar, R. (2024). Genomic insights into fish pathogenic bacteria: A systems approach. *ScienceDirect*.
- Teixeira, R. R., et al. (2021). Towards precision aquaculture: A high performance, cost-effective IoT approach.
- Yang, X., Zhang, S., Liu, J., Gao, Q., Dong, S., & Zhou, C. (2020). Deep learning for smart fish farming: applications, opportunities and challenges.
- Zhang, J., et al. (2025). Review on Omics Approaches in Aquatic Animal Nutrition. *ScienceDirect*.
- Aquaculture Research Institute. (n.d.). Aquaculture Research Institute programs. *University of Idaho*.
- Biofloc Technology. (2020). *Wikipedia*.
- Anastasiadi, D., & Piferrer, F. (2019). Epimutations in developmental genes underlie the onset of domestication in fish. *Molecular Ecology*, 28(11), 2581–2594.
- Føre, M., Frank, K., Norton, T., Svendsen, E., Alfredsen, J. A., Dempster, T., ... Berckmans, D. (2018). Precision fish farming: A new framework to

improve production in aquaculture. *Biosystems Engineering*, 173, 176–193.

Gjedrem, T., & Rye, M. (2018). Selection response in fish and shellfish: A review. *Reviews in Aquaculture*, 10(1), 168–179.

Gui, J. F., & Zhu, Z. Y. (2021). Molecular basis and biotechnological manipulation of sexual determination and reproduction in fish. *Science China Life Sciences*, 64(5), 869–887.

Hoseini, S. M., Yousefi, M., Hoseinifar, S. H., & Van Doan, H. (2019). Effects of dietary herbal antioxidants on growth, immunity, and oxidative stress in aquaculture: A review. *Aquaculture*, 507, 112–123.

Houston, R. D., & Macqueen, D. J. (2020). Atlantic salmon (*Salmo salar* L.) genetics in the 21st century: Taking leaps forward in aquaculture and biological research. *Animal Genetics*, 51(5), 695–705.

Kemski, M., Vargason, T., & Li, C. (2020). Nutrigenomics in aquaculture: Implications for sustainable feed development. *Frontiers in Marine Science*, 7, 703.

Palaiokostas, C. (2021). Genomic selection in aquaculture: Current status and future directions. *Animal Frontiers*, 11(2), 62–69.

Su, Y., Li, M., Li, H., & Chen, J. (2021). Transcriptomic insights into fish immune responses to pathogens. *Fish & Shellfish Immunology*, 112, 78–87. <https://doi.org/10.1016/j.fsi.2021.03.012>

Yáñez, J. M., Yoshida, G. M., & Barría, A. (2020). Integrating genomics into aquaculture to improve disease resistance. *Aquaculture*, 530, 735732.